

# LC-MS/MS ANALYSIS OF IMMUNOSUPPRESSANT DRUGS IN WHOLE BLOOD USING THE XEVO TQ ABSOLUTE WITH CAPITAINER® B 50 DEVICES FOR CLINICAL RESEARCH

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## INTRODUCTION

Traditional laboratory analysis of the immunosuppressant drugs cyclosporine, everolimus, sirolimus and tacrolimus is well-established in clinical research. However there remains a need for individuals to undergo an invasive, time-consuming and disruptive process under the supervision of trained staff in order to collect a sufficient volume of whole blood for laboratory analysis.

A reliable, remote sampling method may find utility in a clinical research setting. Here we describe the use of Capitainer® B 50 Devices to obtain analytically sensitive, precise and accurate data for cyclosporine, everolimus, sirolimus and tacrolimus analysis using small sample volumes for clinical research studies.

The Waters™ ACQUITY™ UPLC™ I-Class FL with Xevo™ TQ Absolute mass spectrometer was used to analyze these samples.

## METHODS

### Materials and Sample Preparation

- MassTrak Immunosuppressant Calibrator and Control Sets were used.
- 30 µL of whole blood was pipetted onto the inlet of the Capitainer® B 50 device, which resulted in a 10 µL dried blood spot.
- Following overnight drying, the dried blood spot was removed and placed in a 2mL microcentrifuge tube.
- 200 µL of internal standard (12.5 ng/mL <sup>2</sup>H<sub>12</sub>-cyclosporine, 1 ng/mL ascromycin, <sup>13</sup>C<sub>2</sub><sup>2</sup>H<sub>4</sub>-everolimus and <sup>2</sup>H<sub>3</sub>-sirolimus in 10% methanol) was added, and the tube underwent mixing and sonication steps.
- Add 10 µL of 0.05M hydrochloric acid and 1 mL *tert*-methyl butyl ether, vortex mix and centrifuge.
- 850 µL of the top layer was transferred to a clean, TruView Total Recovery vial (p/n: 186005669CV), and dried under nitrogen at 40°C.
- Samples were reconstituted in 200 µL mobile phase A:mobile phase B 50:50 (v:v).

### LC-MS/MS Parameters

- Using an ACQUITY UPLC I-Class FL System, samples were injected onto an ACQUITY UPLC HSS C<sub>18</sub> SB Column, 1.8µm 2.1x30mm (p/n: 186004117), using a water/acetonitrile/ammonium fluoride gradient (Table 1) and analyzed with a Xevo TQ Absolute Mass Spectrometer (Figure 1) in ESI+, using MRM parameters described in Table 2.
- The run time is 1.5 minutes (approximately 2.2 minutes injection-to-injection).

Time [min]	Flow rate [mL/min]	A [%]	B [%]	Curve
Initial	0.8	50	50	Initial
0.2	0.8	50	50	1
0.6	0.8	0	100	6
1.2	0.8	50	50	11



Table 1. LC gradient for analysis of the immunosuppressants using the ACQUITY UPLC I-Class FL System

Analyte	Parent (m/z)	Daughter (m/z)	Dwell (s)	Cone (V)	Collision Energy (V)
Cyclosporine	1219.8	1202.8 (1184.8)	0.02	50	18 (34)
<sup>2</sup> H <sub>12</sub> -Cyclosporine	1231.8	1214.8	0.02	50	18
Everolimus	975.6	908.6 (926.6)	0.02	50	16 (12)
<sup>13</sup> C <sub>2</sub> <sup>2</sup> H <sub>4</sub> -Everolimus	981.6	914.6	0.02	50	16
Sirolimus	931.6	864.5 (882.5)	0.02	50	16 (12)
<sup>2</sup> H <sub>3</sub> -Sirolimus	934.6	864.5	0.02	50	16
Tacrolimus	821.5	768.5 (786.5)	0.02	50	20 (18)
Ascromycin	809.5	756.5	0.02	50	22

Table 2. MRM transitions and parameters of the immunosuppressants and internal standards (qualifier parameters in parentheses)



Figure 1. The Xevo TQ Absolute Mass Spectrometer

## RESULTS

Five analytical runs were performed using this method.

### Chromatography and Analytical Sensitivity

- The analytical sensitivity of the developed method for the lowest calibrator (1 ng/mL for everolimus, sirolimus and tacrolimus; 25 ng/mL for cyclosporine) is shown in Figure 2, with S/N (PtP) > 10 for the four analytes over the five runs.

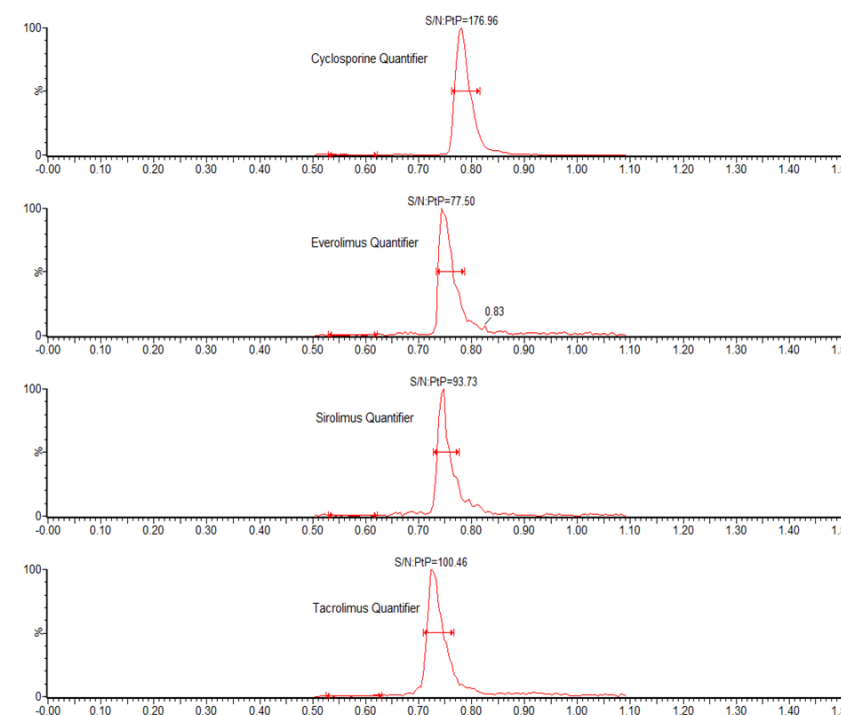


Figure 2. Representative chromatograms of the lowest immunosuppressant calibrator, demonstrating analytical sensitivity

### Linearity and Precision

- Linearity of the calibration ranges (1-30 ng/mL for everolimus, sirolimus and tacrolimus; 25-1500 ng/mL for cyclosporine) was demonstrated with mean  $r^2$  values for the calibration lines >0.99 over five analytical runs.
- Total reproducibility and repeatability across the immunosuppressants at the QC three concentrations (2, 8 and 22 ng/mL for all analytes except cyclosporine, which were 150, 400 and 900 ng/mL), with five\* replicates over five analytical runs (n = 25\*) was ≤7.6%CV (Figure 3).

\*except cyclosporine, four runs and n=20

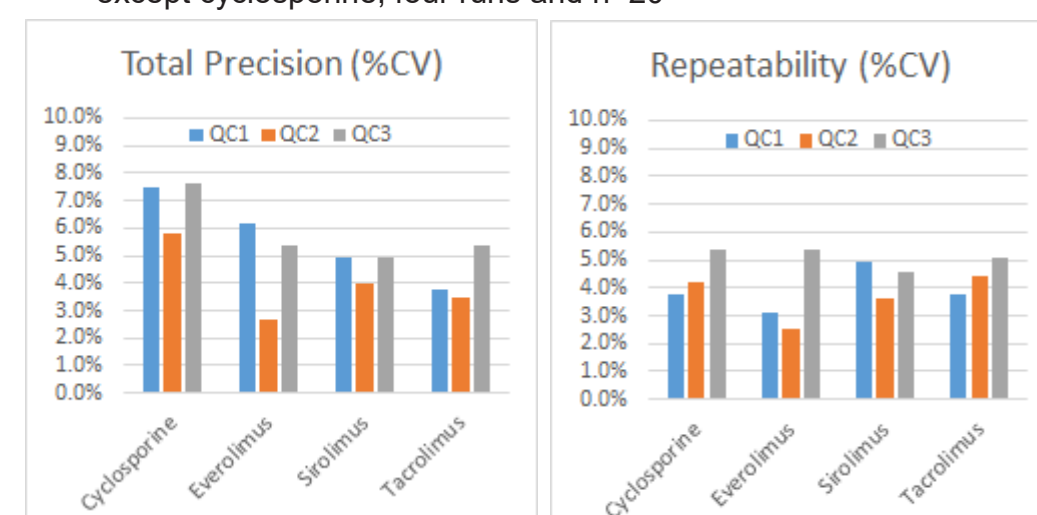


Figure 3. Total reproducibility and repeatability

### Accuracy

- LGC (Bury, UK) whole blood External Quality Assurance samples were sourced and analyzed to assess method accuracy. A summary is presented in Table 3.
- All individual samples met the criteria of the scheme, with mean bias ≤8.7%.

Analyte	Number of samples analyzed (n)	Range (ng/mL)	Mean %bias from Scheme LC-MS ALTM
Cyclosporine	20	31.0-1814.1	+6.0
Everolimus	20	0-23.2	+2.9
Sirolimus	25	1.9-22.9	-8.7
Tacrolimus	25	1.6-27.0	+5.6

Table 3. EQA accuracy summary (note: ALTM is the all-laboratory trimmed mean)

## CONCLUSION

- Capitainer B 50 devices prepared dried blood spots from a low initial whole blood volume, enabling simultaneous analysis of the immunosuppressant drugs cyclosporine, everolimus, sirolimus and tacrolimus in 2.2 minutes (injection-to-injection).
- The performance characteristics of the method indicate good analytical sensitivity, total precision and repeatability (≤7.6%CV) across all analytes and concentrations tested.
- Finally, good agreement was obtained when analyzing External Quality Assurance samples, providing confidence in the accuracy of the method and sample collection devices.

## ACKNOWLEDGEMENT

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Note: ACQUITY™, UPLC™ and Xevo™ are trademarks owned by Waters Technologies Corporation. Capitainer B is a registered trademark of Capitainer AB.